

## THE EFFECT OF LIPOPOLYSACCHARIDE ON INTERLEUKIN-1A PRODUCTION BY PERIPHERAL BLOOD LEUKOCYTES *IN VITRO* AT VARIABLE TIMES AND TEMPERATURES.

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### Abstract

The concentration of Interleukin-1 $\alpha$  (IL-1 $\alpha$ ) was measured in peripheral blood leukocytes (PBLs) culture at variable times (0, 20 h, 23 h, 43 h, and 45 h) in presence or absence of lipopolysaccharide (LPS) at variable temperatures (35 °C, 37 °C and 40 °C). There was not high difference in IL-1 $\alpha$  concentrations at 35°C and 40°C in compared with control group (37 °C) and the same results were found in cultures that supplemented with LPS. There was a slight elevated (Non significant elevated) in IL-1 $\alpha$  concentrations at 35 °C + LPS after 45h and slight decrease at 40 °C+LPS after 43h in compared with control (37 °C+LPS), while significant increases were found in cultures supplemented with LPS at variable times and at variable temperatures in compared with opposites temperatures and times with cultures that did not supplement with LPS.

### Introduction

Interleukins-1 are classical multifunctional cytokines, they are produced by macrophage, other antigen presenting cells (APCs), most of somatic cells, T-cell and activated B cell that products Immunoglobulin. Interleukins-1 activates phagocyte and promotes of hematopoiesis (1,2). The IL-1 family comprises IL-1 $\alpha$  and IL-1 $\beta$  as well as the IL-1 receptor antagonist (IL-1Ra), two types of IL-1 receptors have been identified in human. Although IL-1 $\alpha$  and IL-1 $\beta$  share little homology in primary sequence the two cytokines exert their against effects through binding to the same type I receptor (IL-1 RI), type II receptor (IL-1 RII) also binds IL-1 but does not transducer signal (1,3). The receptor antagonist (IL-1 Ra) binds to IL-1 RI with an affinity comparable to that of IL-1 $\alpha$  and IL-1 $\beta$ , but does not elicit biological responses (4,5). Furthermore, intracellular fragments of both receptor types (IL-1 RI and IL-1 RII) circulate in the blood and function as natural scavengers by binding and neutralizing IL-1 $\alpha$ , IL-1 $\beta$  or IL-1 Ra (6). The complex array of interacting factor could find modulate IL-1 $\alpha$  responses in vitro (7). Lipopolysaccharide (LPS) recognition by higher animals involves soluble protein (LPS-binding protein [LBP] and soluble CD<sub>14</sub> [sCD<sub>14</sub>]) (8). Membrane receptors (CD<sub>14</sub> and Toll-like receptor 4 (TLR4)) (8). LBP promotes rapid binding of purified LPS

from aggregates to membrane-bound CD<sub>14</sub> (mCD<sub>14</sub>) on cells or to sCD<sub>14</sub> in plasma (9). CD<sub>14</sub> is important for conferring sensitive cellular responses to LPS and TLR4 appears to be the most important LPS signal transducer (10, 11). Bacterial LPS can activates a variety of mammalian cell types (part of them PBPLs) and is a powerful activator of the innate immune system, LPS stimulates the synthesis and release of proinflammatory cytokines such as IL-1 from monocytes and macrophages, these cytokines can further activate monocytes, neutrophils and lymphocytes, initiating cellular injury and tissue damage (12,13). LPS produces fever after 60- 90 minutes that occur after stimulation of IL-1 production from APCs and this carries by the blood stream to the thermoregulatory center in the hypothalamus where physiologic responses are initiated that result in fever and this phenomenon (fever) play an important role in the first step of immune response, so some beneficial effects of fever on the control of infection in a few instances for example antibody production and T-cells proliferation are more efficient at high body temperature (14,15,16). There is proof that IL-1 $\alpha$  can modulate LPS activity and its effect in human body (17, 18). The aim of this study was conducted to evaluate the effect of LPS and temperature alone or together on IL-1 $\alpha$  production.

## Material and methods:

1- Preparation of human peripheral blood leukocytes (PBLs): Peripheral blood buffy coats from healthy donors (five donors) were centrifuged at 2000 rpm (500 x g) for 5 min. Erythrocytes in the pellet were lysed in sterile 0.83 % NH<sub>4</sub> Cl solution for 3 min. at room temperature and PBLs were washed twice by centrifugation (500 x g for 5 min.) in phosphate buffered saline (PBS : 137 mM NaCl , 10 mM Na<sub>2</sub>HPO<sub>4</sub> , 2.7 mM KCl and 1.8 mM KH<sub>2</sub>PO<sub>4</sub> [ PH : 7.23 } ) (19). The viability of PBLs was measured by trypan blue dye exclusion test (20) and number of PBLs was adjusted to 10<sup>6</sup> viable cells / ml with culture medium.

2- Preparation of culture medium: The medium that used was RPMI 1640 (Sigma) supplemented with 10% fetal bovine serum, 20 mM HEPES and 2 mM L- glutamine.

3- Treatment of cells : Every donor PBLs solution divided into many test tubes (sterile) and these test tubes divided into two groups. Group A, and group B first group supplemented with 1μ/ml of LPS ( O 127 : Bs Difco, Betroit Mich.) and group B did not supplemented with LPS . Every group divided into three subgroup of tubes. The first subgroups incubated at 35 °C , second subgroups incubated at 37 °C and third subgroups incubated at 40°C (each tube was contained 1 ml of medium that containing 10<sup>6</sup> PBLs) after that the supernatants were collected from all tubes by centrifugation at 500 x g /10 min. after (Zero time , 20 h., 23 h., 43h., and 45 h.), the supernatants were stored at -20 in microcentrifuge tubes.

4- Detection of IL-1α: IL-1α level was measured by using human cytokine immunoassay kit (Bachman coulter, Marseille Cedex 9, France) according to the manufacturer's protocol.

5- Statistical analysis: t – test was performed.

## Results and discussion

The concentrations of IL-1α were detected in cultures of PBLs ,slight significant increase was found in this concentration at 35 °C but at 23, 43h and

45h in compared with (control) for all times .At 40°C slight decreases were found in the concentrations of IL-1α at 43h and 45h only when compared these results with control (37 °C for all times) Table (1) .

**Table (1)**

**The concentration of IL-1α pg/ml at variable times and at variable temperatures**

Temperatures	20 h	23 h	43 h	45 h
35 °C	128.7 ± 8.5 NS	159 ± 9.5 P<0.05	130 ± 7.9 P<0.05	125 ±10.4 P<0.05
37 °C control	130 ± 7.6	140 ± 8.3	104 ± 10.1	110 ±9.7
40 °C	139 ±12. 4 NS	130 ± 11.9 NS	88 ± 9.6 P<0.05	93 ± 10.1 P<0.05

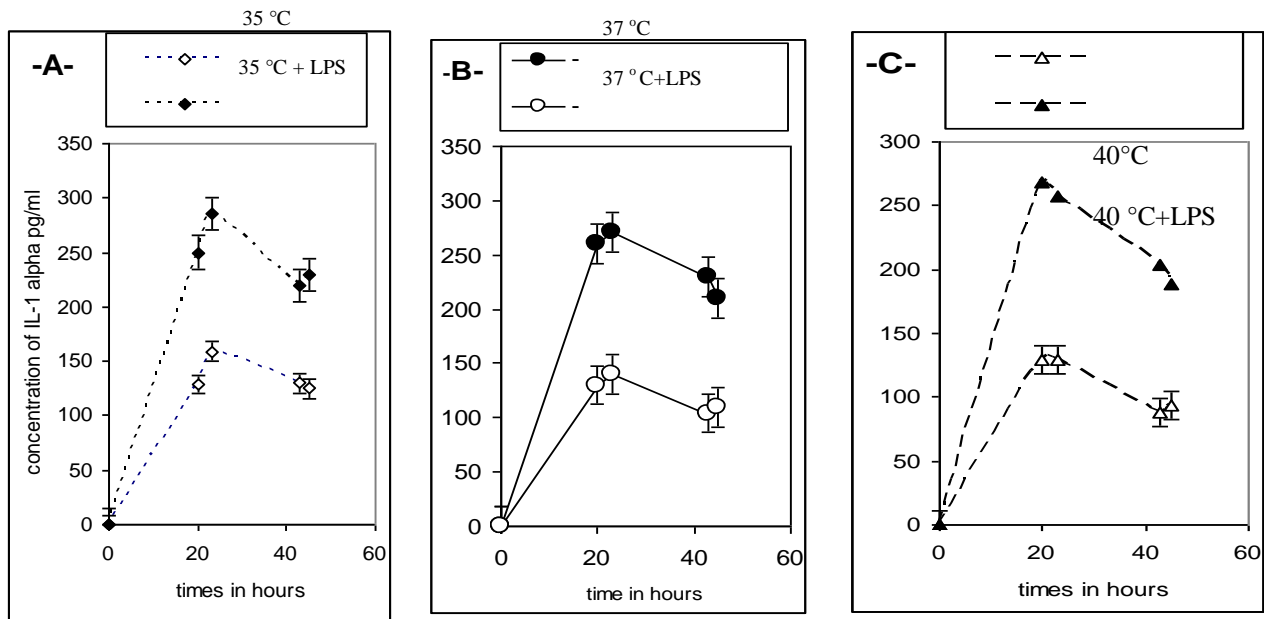
In compared the concentrations of IL-1α in (35 °C+LPS and 40°C+LPS) with control (37°C+LPS) the slight different was found .There is little increase in concentration of IL-1α at ( 35°C+LPS) at 45h only , and decrease in concentration of IL-1α at(40+LPS) at 43h and 45h when compared with control (37°C+LPS) Table (2) .

**Table (2)**

**The concentration of IL-1α pg/ml at variable times and at variable temperatures in vitro in presence of LPS.**

Temperatures	after 20 h	after 23h	after 43h	after 45h
35 °C+ LPS	251 ± 15.5 NS	286 ± 14.1 NS	220 ±16.2 NS	230 ± 13.4 P<0.05
37 °C+LPS ( control)	260 ± 19.7	271 ± 18.3	230 ± 15.9	210 ± 17.8
40 °C + LPS	268 ± 17.1 NS	258 ± 18.2 NS	204 ± 20.2 p<0.05	188 ± 19.4 P<0.05

When compared the concentration of IL-1α between presence and absence of LPS in all temperatures for all times. The concentrations of IL-1α in presence of LPS were higher than the concentration of IL-1α in absence of LPS in every temperatures and all times Figure (1).



**Figure 1- Comparison of concentrations of IL-1 $\alpha$  in presence and absence of LPS in many temperatures: 1- (A) 35 °C, 2- (B) 37 °C and 3- (C) 40 °C**

LPS potently stimulates the production of IL-1 and which has a large number of biological activities, thus activates of LPS are believed to be mostly mediated by IL-1 or cytokine network involving IL-1 (2, 21, 22). Other investigators stimulated cells to produce IL-1 in vitro; they found significant increase in IL-1 production at 3 and 20 h of incubation (23), these results that obtained by last investigator sometimes agree with our results. We documented LPS could stimulate PBLs to production of IL-1 $\alpha$  at every temperatures more than cultures that were not supplemented with LPS. LPS play an important role in activation of normal cells to produce IL-1 and other types of cytokines by binding (LPS) with special receptors on these cells. This binding will reach activated signal to the NF $\kappa$ B and this will activate special gene that responsible for production of IL-1 and other proinflammatory cytokines thus LPS has high ability to produce inflammation in human body and animals (8).

IL-1 produces by many types of normal cells besides PBLs such as fibroblast, endothelial cells epithelial cells and other types of cells (2, 24). Besides normal cells tumor cells can produce IL-1 and last play a crucial role in regulation of immune system against tumor in special places (25). IL-1 may produces

after stimulation with foreign stimulator or by self stimulator such as other cytokines or produce by autocrine stimulation (cytokine stimulates its cell to produce the same cytokine) (26), variable temperatures have ability to stimulate of immune system (14, 15). But we did not find any evidence of the effect of temperatures on IL-1 production. but Interleukines-1 are an important inducer of numbers of effects that are mediated through hypothalamus, they are endogenous pyrogens (they induce fever) (1) that is meaning IL-1 elevates temperature but the last do not affect on IL-1 and we suggest there is not any synergistic effects between temperatures and LPS on IL-1 production by peripheral blood leukocytes in vitro.

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## الخلاصة

تم حساب تركيز الأنترلوكين-1 من نوع الفا (IL-1 $\alpha$ ) في المزارع الخاصة بكريات الدم البيض المأخوذة من الدم المحيطي لعدة اشخاص اصحاء ولفترات زمنية مختلفة ( صفر , 20 ساعة , 23 ساعة , 43 ساعة و 45 ساعة ) بوجود او عدم وجود عديد السكريد الشحمي (LPS) وبدرجات حرارة مختلفة ( 35 م<sup>°</sup> , 37 م<sup>°</sup> و 40 م<sup>°</sup> ) حيث لم يلحظ تغيرات واضحة وكبيرة في تركيز الأنترلوكين-1 الفا في درجة حرارة 35 م<sup>°</sup> و 40 م<sup>°</sup> عند مقارنة النتائج مع مجموعة السيطرة اي بدرجة حرارة 37 م<sup>°</sup> . اما بوجود LPS كذلك لم نلاحظ الأتغير طفيف اي ارتفاع بسيط في تركيز (IL-1 $\alpha$ ) في درجة حرارة 35 م<sup>°</sup> وبعد 45 ساعة . كذلك لوحظ انخفاض بسيط في هذا التركيز عند درجة حرارة 40 م<sup>°</sup> ولكن بعد 43 ساعة بالمقارنة مع مجموعة السيطرة (37 درجة مئوية + LPS) . بينما لوحظ ارتفاع معنوي في تركيز IL-1 في المزارع المضاف اليها LPS وعند جميع درجات الحرارة وفي جميع الاوقات بالمقارنة مع المزارع التي لم يستخدم فيها LPS وكذلك في جميع درجات الحرارة و الاوقات المناصرة لها .